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### The nature of a supposed $N_2$ complex of ferroleghaemoglobin

ABEL AND BAUER<sup>1</sup> prepared leghaemoglobin (Lb) by grinding soybean root nodules in 3 M  $(NH_4)_2SO_4$  at pH 9 under  $H_2$  followed by fractionation of the crude extract with excess  $(NH_4)_2SO_4$ . This ferrous leghaemoglobin ( $Lb^{2+}$ ) was claimed to be isolated as a  $Lb^{2+}N_2$  complex<sup>2</sup> since addition of  $Na_2S_2O_4$  did not cause the disappearance of specific absorption peaks thought to be due to a  $Lb^{2+}$ -gas complex. If the gas had been  $O_2$  the spectrum should have reverted to that of  $Lb^{2+}$  (cf. Fig. 1, dashed trace). The addition of  $K_3Fe(CN)_6$  to their preparation resulted in the liberation of gas following oxidation of the haem iron; the gas was claimed to be identified as  $N_2$  by a gas-chromatographic procedure<sup>2</sup>.

When their preparative procedure was repeated in our laboratory using nodules picked from "Lincoln" strain soybean roots inoculated with *Rhizobium japonicum* strain 505 (Wisconsin)<sup>3</sup> the final precipitate, redissolved in 0.1 M potassium phosphate (pH 6.8) or 0.1 M Tris (pH 7.9) showed absorption peaks at 538 and 574 nm, characteristic of  $Lb^{2+}O_2$ , and a small shoulder at 626–628 nm, characteristic of oxidized leghaemoglobin ( $Lb^{3+}$ ). Titration of such preparations with  $Na_2S_2O_4$  caused the collapse of these peaks and shoulder and in the final spectrum (Fig. 1, solid trace) the broad peak at 558 nm was consistent with the formation of  $Lb^{2+}$  and the 550-nm peak and 520-nm shoulder were assumed to be due to the " $Lb^{2+}N_2$  complex" for which no spectrum had previously been presented<sup>2</sup>.

1.06  $\mu$ moles of  $Lb^{2+}$  containing this supposed " $Lb^{2+}N_2$  complex", diluted to 3 ml in 0.1 M phosphate (pH 6.8), were placed in a 10-mm light-path optical cuvette

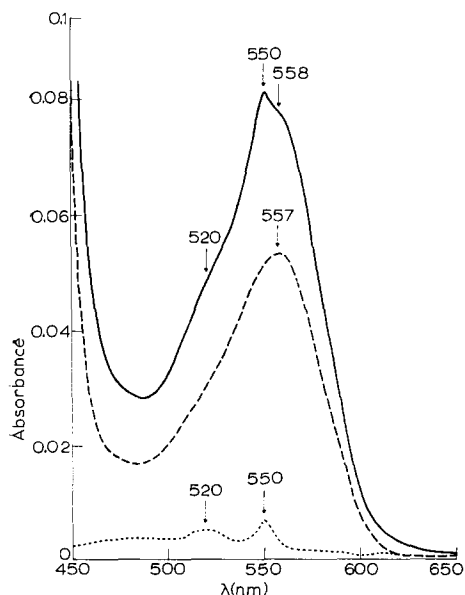


Fig. 1. Spectra of the supposed " $Lb^{2+}N_2$  complex" and its separated components, measured in a Cary 14 spectrophotometer using 10-mm light-path cuvettes. All samples reduced with  $Na_2S_2O_4$  (1 mg/ml). Experimental details are given in the text. —, " $Lb^{2+}N_2$ ", before chromatography; ---,  $Lb^{2+}$ , after chromatography; ..... reduced cytochrome *c* after chromatography.

fused to the bottom of a Thunberg tube, and gassed with pure  $N_2$  to allow complete formation of any  $N_2$  complex, followed by evacuation and refilling with pure argon to 0.1 atm. Spectrophotometric analyses<sup>4</sup> showed that the  $Lb^{2+}$  in this solution had become about 50% oxygenated ( $0.5 \mu\text{mole } Lb^{2+}O_2$ ) during these manipulations, and that the solution contained about 5% of  $Lb^{3+}$ . This Thunberg tube was connected through an attached tap and B12 "Quickfit" glass socket to the sample inlet system of an Atlas M-86 mass spectrometer. Analysis of the gas phase (17 ml) before and after conversion of all  $Lb$  to  $Lb^{3+}$  by  $6 \mu\text{moles of } K_3Fe(CN)_6$  added from the Thunberg tube sidearm showed that  $0.63 \mu\text{mole } O_2$ , but less than  $0.08 \mu\text{mole } N_2$  were liberated.

A similar preparation of this supposed " $Lb^{2+}N_2$  complex" was dialysed against 1 mM phosphate (pH 6.8) and passed through a small column of Whatman cellulose phosphate powder, equilibrated with 1 mM phosphate (pH 6.8). The first, major effluent fraction had an absorption spectrum characteristic of conventional  $Lb^{2+}$ , with 557-nm peak (Fig. 1, dashed trace). A second, minor band of coloured material, eluted with 1 mM phosphate-100 mM NaCl (pH 6.8), had the absorption spectrum characteristic of reduced cytochrome *c* (Fig. 1, dotted trace) with 550- and 520-nm absorption peaks.

Crude cytochrome *c* (550, Rhizobium) was liberated from conventionally isolated nodule bacteria (Rhizobium bacteroids)<sup>3</sup> following re-extraction with 3.0 M  $(NH_4)_2SO_4$  at pH 9. After purification on ion-exchange columns<sup>5</sup> a mixture of this homogeneous reduced cytochrome *c* (1-2%) with 98-99% of  $Lb^{2+}$  had the spectral properties of the " $Lb^{2+}N_2$  complex" (Fig. 1, solid trace).

It is suggested that the " $Lb^{2+}N_2$  complex" apparently detected<sup>2</sup> by spectrophotometry of  $Na_2S_2O_4$ -reduced, alkaline-extracted  $Lb$  is in fact a simple physical mixture of reduced cytochrome *c* and  $Lb^{2+}$ . The gas liberated by oxidizing this "complex" in the absence of excess  $Na_2S_2O_4$  appears to be  $O_2$  rather than  $N_2$ . These results, and observations to be reported elsewhere on the state of  $Lb$  *in vivo*, do not support a role for  $Lb$  in  $N_2$  transport or activation during symbiotic  $N_2$  fixation.

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